

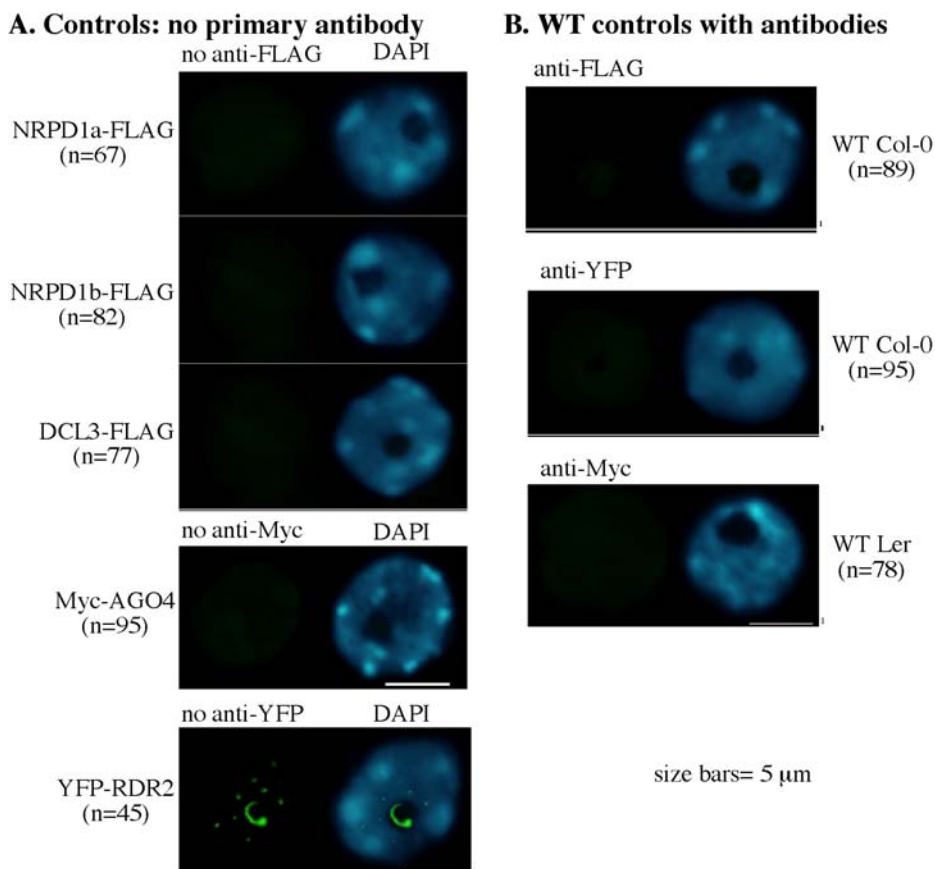
## Supplemental Data

### The *Arabidopsis* Chromatin-Modifying

### Nuclear siRNA Pathway Involves

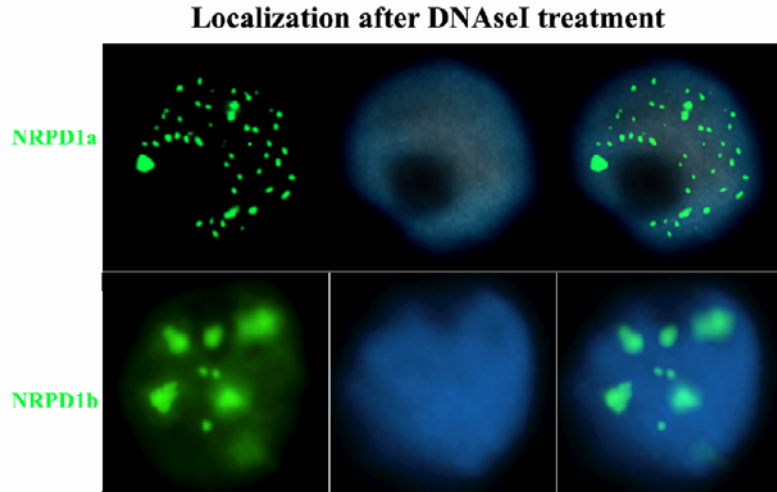
### a Nucleolar RNA Processing Center

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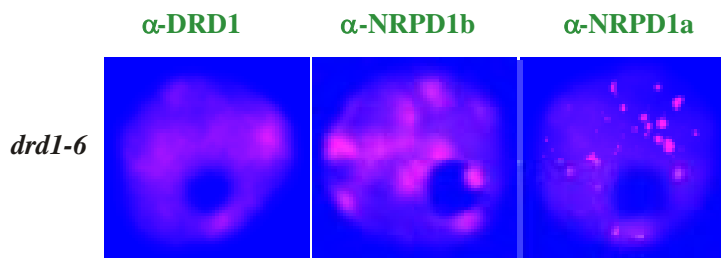
**Figure S1. Antibody Specificity Controls**

In part A of the figure, nuclei of transgenic lines expressing the indicated epitope tagged proteins were processed for protein immunolocalization as in Figure 3 of the paper except that the primary antibody was omitted prior to incubation with FITC-labeled secondary antibody (green). YFP fluorescence accounts for the YFP-RDR2 signal in the absence of anti-YFP antibody. In part B, non-transgenic, wild-type *A. thaliana* (ecotypes Col-0 or Ler) controls show that no signals are obtained upon immunolocalization using anti-FLAG, anti-YFP or anti-Myc primary antibodies. The images shown are representative of the nuclei observed, with the total number analyzed shown in parentheses. Nuclei were counterstained with DAPI (blue); the size bar corresponds to 5  $\mu$ m.



**Figure S2. NRPD1a and NRPD1b Immunolocalization Signals Are Not Lost in DNase I-Treated Nuclei**

Native NRPD1a and NRPD1b proteins were localized using anti-peptide antibodies in nuclei treated with DNase I as described in Figure 3B of the main paper.



**Figure S3. Immunolocalization of DRD1, NRPD1b, and NRPD1a in *drd1-6* Mutant Nuclei**

Proteins were detected using anti-peptide antibodies. Note that DRD1 is not detected in the mutant, suggesting that the antibody specifically recognizes DRD1. The *drd1-6* mutation typically does not affect the NRPD1a pattern (85% yield the wild-type pattern for NRPD1a shown below;  $n = 90$ ) but NRPD1b immunolocalization signals are typically more diffuse in *drd1-6* (79%;  $n = 79$ ) than in wild-type, suggesting that DRD1 may act upstream, or at the same step, as NRPD1b.

**Table S1. Supporting Data for Figure 2A: siRNA Probe Hybridization Patterns and Frequencies**

RNA probe	Localization phenotypes	Frequency (%) of phenotypes observed upon nuclease treatment or in different genetic backgrounds									
		Col	Ler	+RNase A	+DNase I	<i>nrdp1a</i>	<i>nrdp2</i>	<i>nrdp1b</i>	<i>rdr2-1</i>	<i>dcl3-1</i>	<i>ago4-1</i>
45S siR	Nucleolar dot observed:	100	100	0	100	0	0	0	0	0	0
	Dispersed nuclear signal:	0	0	0	0	29	13	56	8	9	29
	No signal:	0	0	100	0	71	87	44	92	91	71
	# nuclei observed	n = 75	n = 71	n = 71	n = 63	n = 65	n = 141	n = 132	n = 62	n = 72	n = 76
5S siR	Nucleolar dot observed:	100	100	0	100	0	0	0	0	0	0
	Dispersed nuclear signal:	0	0	0	0	0	6	75	11	3	17
	No signal:	0	0	100	0	100	94	25	89	97	83
	# nuclei observed	n = 56	n = 48	n = 62	n = 68	n = 81	n = 127	n = 162	n = 85	n = 62	n = 74
AtSN1	Nucleolar dot + nucleoplasm:	74	No data	0	89	No data	0	No data	0	No data	No data
	Nucleoplasm only:	26		0	11		0		0		
	No signal:	0		100	0		100		100		
	# nuclei observed	n = 67		n = 79	n = 85		n = 150		n = 123		
AtCopia4	Nucleolar dot + nucleoplasm:	100	No data	0	100	No data	0	No data	0	No data	No data
	No signal:	0		100	0		100		100		
	# nuclei observed	n = 85		n = 53	n = 68		n = 103		n = 91		
45S precursor	Diffuse nucleolar signals:	100	100	100	100	100	100	100	100	100	100
	# nuclei observed	n = 63	n = 57	n = 64	n = 51	n = 86	n = 79	n = 127	n = 72	n = 74	n = 81

The table is organized as in Figure 2A except that the table includes two columns of data for wild-type nuclei (ecotypes Col-0 and Ler) whereas Figure 2A showed only the Col-0 wild-type control.

**Table S2. Supporting Data for Figure 3A: Protein Localization and Effects of RNase**

	NRPD1a	NRPD2	NRPD1b	RDR2	DCL3	AGO4
<b>protein localization</b>	100% of nuclei display pattern shown  n = 82	100% of nuclei display pattern shown  n = 245	100% of nuclei show the nucleolar dot. 57% display numerous puncta external to nucleolus, as shown; 43% show <10 puncta  n = 77	100% of nuclei display pattern shown  n = 87	100% of nuclei display pattern shown  n = 125	100% of nuclei display pattern shown  n = 96
<b>Effect of RNase A</b>	91% , protein not detectable 9% , WT pattern  n = 85	81% , protein not detectable 19% , WT pattern  n = 94	65% , protein not detectable 35% , WT pattern  n = 93	85% , protein not detectable 15% , WT pattern  n = 62	59% , protein not detectable 41% , WT pattern  n = 89	72% , protein not detectable 28% , WT pattern  n = 61

**Table S3. Supporting Data for Figure 3C: Pairwise Detection of Nuclear siRNA Pathway Proteins**

Epitope-tagged lines	Antibodies				
	$\alpha$ -NRPD1a	$\alpha$ -NRPD2	$\alpha$ -NRPD1b	$\alpha$ -RDR2	$\alpha$ -DCL3
<b>NRPD1a-FLAG</b>		Majority of the nucleoplasmic signals colocalized  n = 93			
<b>NRPD1b-FLAG</b>	Few nucleoplasmic signals colocalized  n = 71	Few nucleoplasmic signals colocalized  n = 85			
<b>YFP-RDR2</b>	Few nucleoplasmic signals colocalized  n = 54	Few nucleoplasmic signals colocalized  n = 48	Nucleolar dot + Few nucleoplasmic signals colocalized  n = 67		
<b>DCL3-FLAG</b>	Few nucleoplasmic signals colocalized  n = 76	Few nucleoplasmic signals colocalized  n = 81	Nucleolar dot + Few nucleoplasmic signals colocalized  n = 73	Nucleolar dot + Few nucleoplasmic signals colocalized  n = 86	
<b>Myc-AGO4</b>	Not colocalized  n = 54	Few nucleoplasmic signals colocalized  n = 61	Nucleolar dot + Few nucleoplasmic signals colocalized  n = 58	Nucleolar dot colocalized  n = 45	Nucleolar dot colocalized  n = 59

**Table S4. Supporting Data for Figure 4: Protein-siRNA Colocalization**

RNA probes	Epitope-tagged lines				
		NRPD1b-Flag	YFP-RDR2	DCL3-Flag	cMyc-AGO4
45S siR	Colocalized	81%	82%	79%	91%
	Not colocalized	19% <i>n</i> = 46	18% <i>n</i> = 60	21% <i>n</i> = 75	9% <i>n</i> = 65
siR1003	Colocalized	76%	58%	85%	76%
	Not colocalized	24% <i>n</i> = 57	42% <i>n</i> = 72	15% <i>n</i> = 79	24% <i>n</i> = 57
AtSN1	Colocalized	85%	61%	76%	83%
	Not colocalized	15% <i>n</i> = 74	39% <i>n</i> = 56	34% <i>n</i> = 45	17% <i>n</i> = 56
AtCopia4	Colocalized	82%	54%	78%	72%
	Not colocalized	18% <i>n</i> = 57	46% <i>n</i> = 59	22% <i>n</i> = 49	28% <i>n</i> = 67
45S prec	Colocalized	25%	43%	21%	30%
	Not colocalized	75% <i>n</i> = 81	57% <i>n</i> = 64	79% <i>n</i> = 61	70% <i>n</i> = 75

Colocalization was considered to be when >50% of the RNA probe signal overlapped >50 % of the protein signal.

**Table S5. Supporting Data for Figure 5: Localization of Proteins Relative to NORs and 5S Gene Loci**

DNA loci		NRPD1a	NRPD2a	NRPD1b	RDR2	DCL3	DRD1
NORs	Colocalized	85%	93%	92%	22%	12%	87%
	Not colocalized	15% <i>n</i> = 71	7% <i>n</i> = 83	8% <i>n</i> = 89	78% <i>n</i> = 55	88% <i>n</i> = 66	13% <i>n</i> = 57
5S gene clusters	Colocalized	68%	72%	81%	13%	27%	72%
	Not colocalized	32% <i>n</i> = 58	28% <i>n</i> = 62	19% <i>n</i> = 76	87% <i>n</i> = 51	73% <i>n</i> = 65	28% <i>n</i> = 61

Colocalization was considered to be when at least two NORs and at least four 5S gene *loci* overlapped half of the protein signals outside the nucleolus.

**Table S6. Supporting Data for Figure 6: Protein Localization in Various Nuclear siRNA Pathway Mutants**

		NRPD2	NRPD1a	NRPD1b	RDR2	DCL3
WT	Col	100% of nuclei display pattern shown <i>n</i> = 245	100% of nuclei display pattern shown <i>n</i> = 160	71% of nuclei display pattern shown <i>n</i> = 185	77% of nuclei display pattern shown <i>n</i> = 96	100% of nuclei display pattern shown <i>n</i> = 125
	<i>nrd1a</i>	Reduction in labeling intensity <i>n</i> = 181	No signal <i>n</i> = 123	WT pattern <i>n</i> = 87	Very faint to no signal <i>n</i> = 145	Very faint to no signal <i>n</i> = 61
Mutants	<i>nrd2</i>	Not detected <i>n</i> = 155	Reduction in labeling intensity <i>n</i> = 178	Very faint to no signal <i>n</i> = 134	Very faint to no signal <i>n</i> = 141	Very faint to no signal <i>n</i> = 104
	<i>nrd1b</i>	Very faint to no signal <i>n</i> = 138	WT pattern <i>n</i> = 67	No signal <i>n</i> = 149	Nucleolar dot is not detected <i>n</i> = 153	- Very strong reduction in labeling intensity (76%) - Mislocalization of the nucleolar dot to the nucleoplasm (24%) <i>n</i> = 84
	<i>nrd2, nrd1a</i>	Very faint to no signal <i>n</i> = 74	Very faint to no signal <i>n</i> = 81	Very faint to no signal <i>n</i> = 90	Very faint to no signal <i>n</i> = 67	Very faint to no signal <i>n</i> = 57
	<i>rdr2-1</i>	Small reduction in labeling intensity <i>n</i> = 121	WT pattern <i>n</i> = 112	- Nucleolar dot not detected (81%) - Reduction in labeling intensity (19%) <i>n</i> = 157	No signal <i>n</i> = 61	Very faint to no signal <i>n</i> = 87
	<i>dcl3-1</i>	Small reduction in labeling intensity <i>n</i> = 130	WT pattern <i>n</i> = 74	- Nucleolar dot not detected (78%) - Reduction in labeling intensity (22%) <i>n</i> = 72	WT pattern <i>n</i> = 89	No signal <i>n</i> = 91
	<i>ago4-1</i>	Small reduction in labeling intensity <i>n</i> = 109	WT pattern <i>n</i> = 65	- Nucleolar dot not detected (92%) - Reduction in labeling intensity (8%) <i>n</i> = 133	WT pattern <i>n</i> = 122	- WT pattern (67%) - Mislocalization of the nucleolar dot to the nucleoplasm (33%) <i>n</i> = 152